



Interactive Visual Analysis of Biological Data

Tutorial: Interactive Visual Analysis of Scientific Data

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Berkeley Drosophila Transcription Network Project

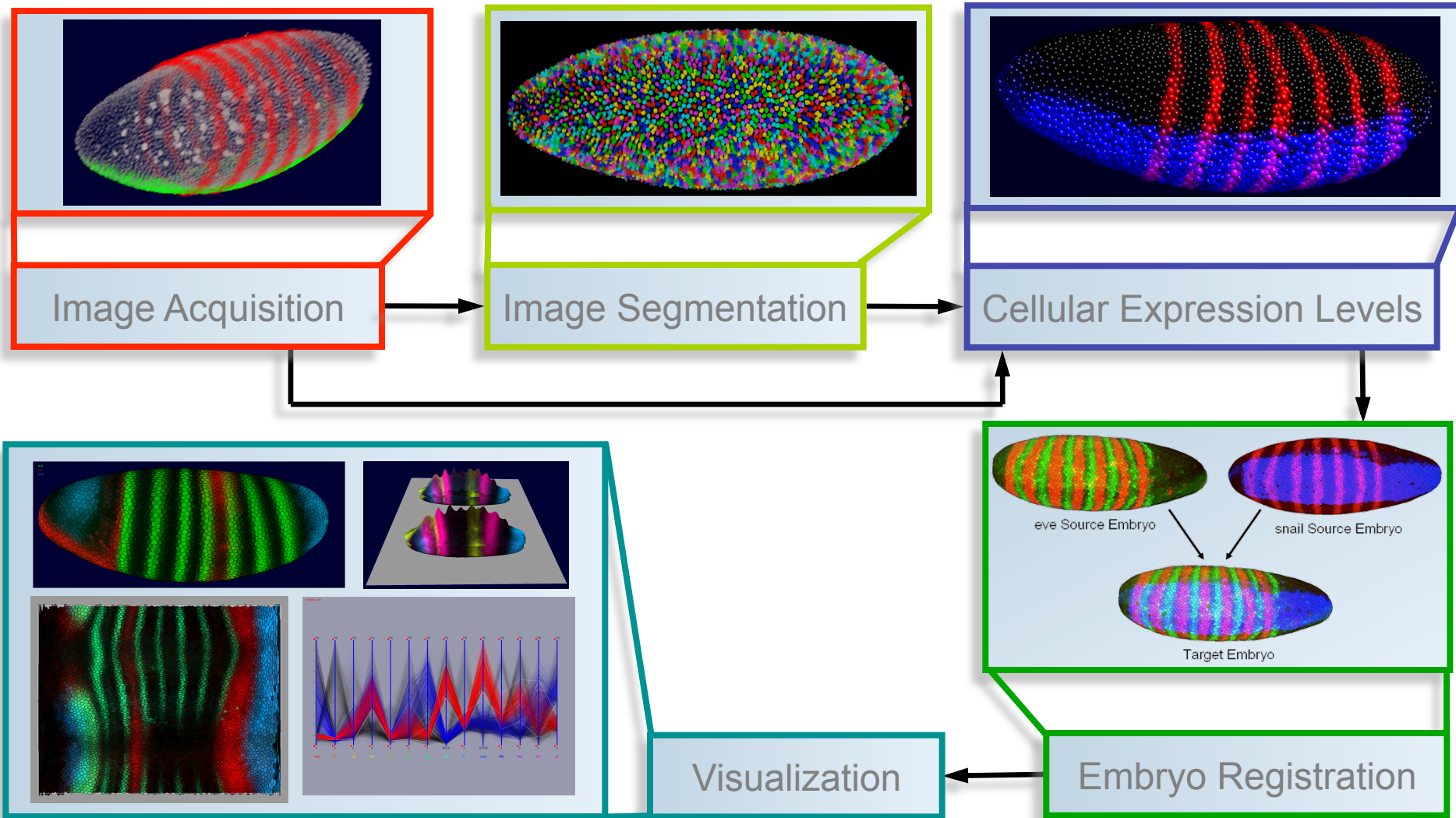


Interactive Exploration of 3D Gene Expression Data

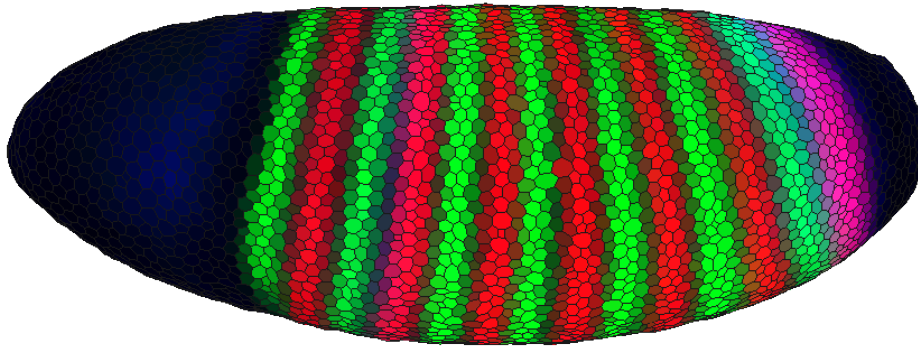
G.H. Weber, O. Rübel, M.-Y. Huang, A. H. DePace, C. C. Fowlkes, S. V. E. Keränen, C. L. Luengo Hendriks, H. Hagen, D. W. Knowles, J. Malik, M. D. Biggin and B. Hamann: *Visual Exploration of Three-dimensional Gene Expression Using Physical Views and Linked Abstract Views* .
IEEE Trans. Comp. Bio. Bioinform., 6(2): 296-309 (2009)



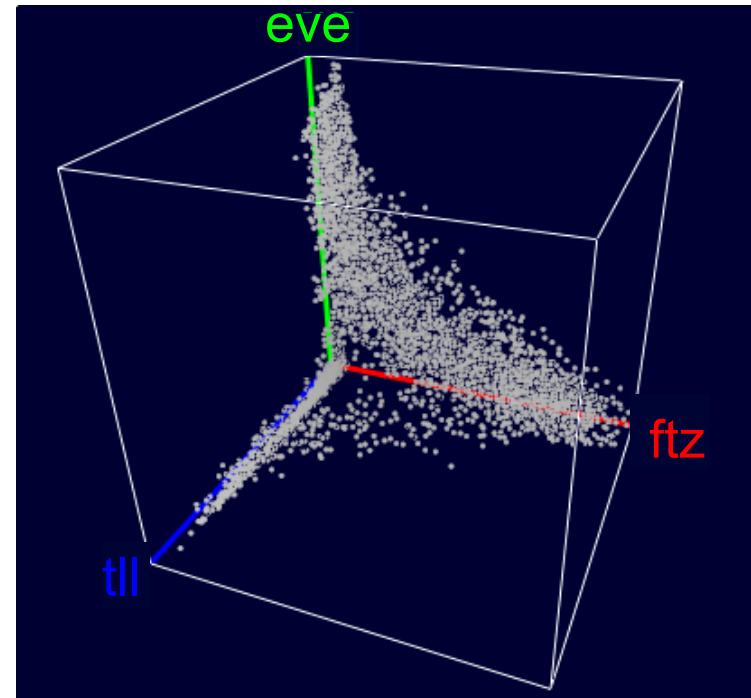
Analysis of 3D Gene Expression Patterns



Multiple Views For Different Aspects of Gene Expression

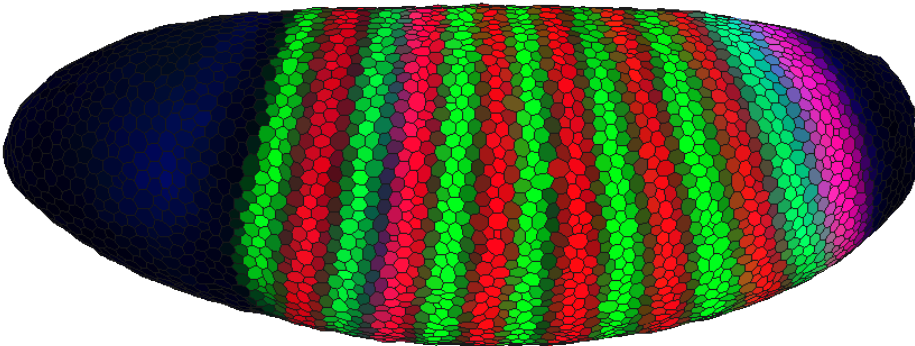


Physical views show gene expression as spatial patterns in relation to the embryo

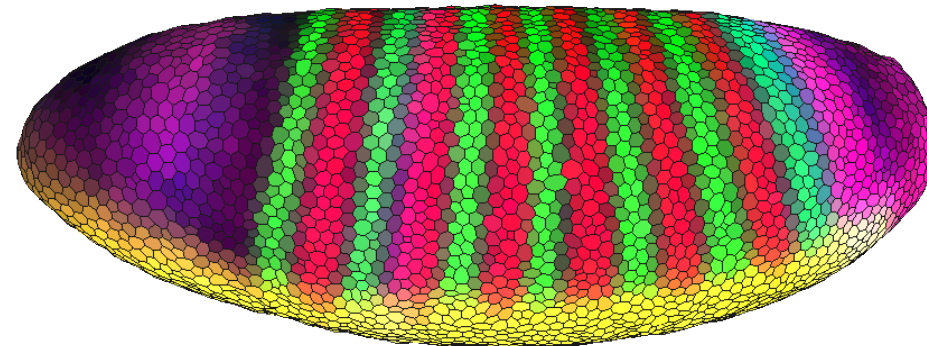


Expression space views show the relationship between expression of different genes

3D Embryo View



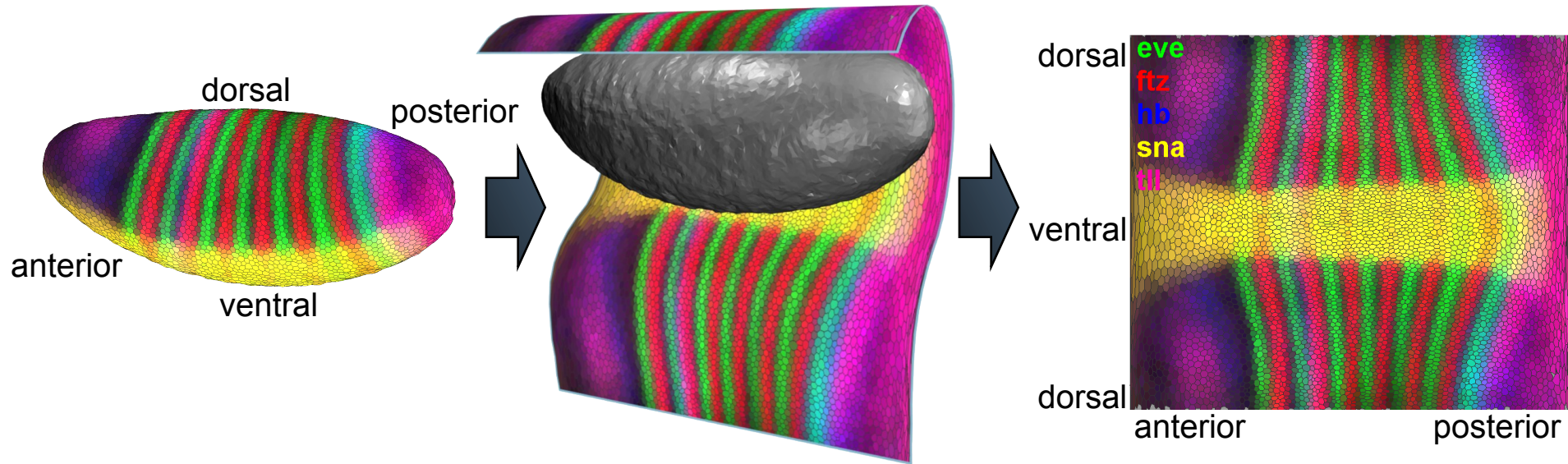
ftz eve hb



ftz eve hb sna tll

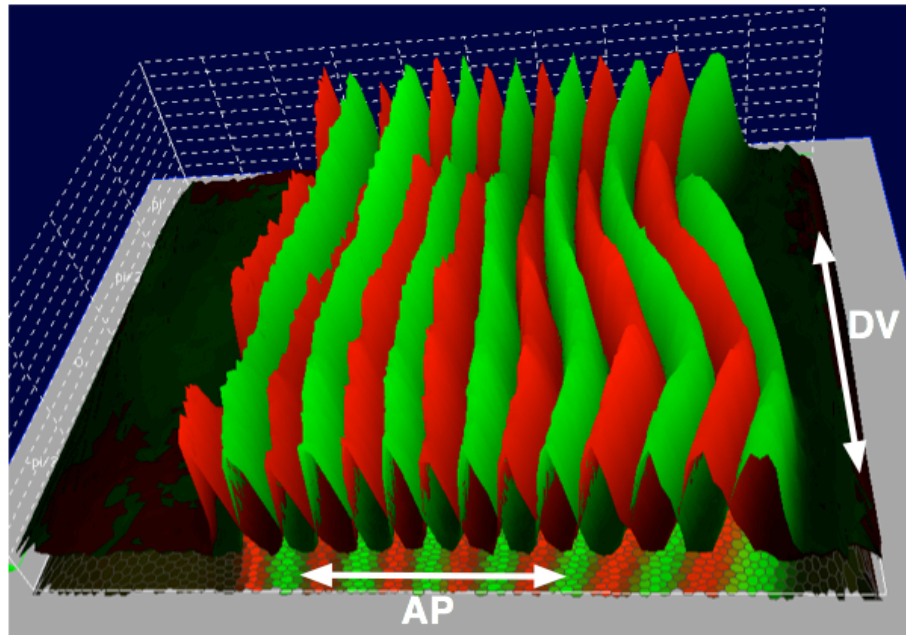
- Each cell represented by a 3D object at the physical cell location
- Usage:
 - Qualitative analysis of gene expression levels
 - Analysis and comparison of gene expression patterns

Unrolled View

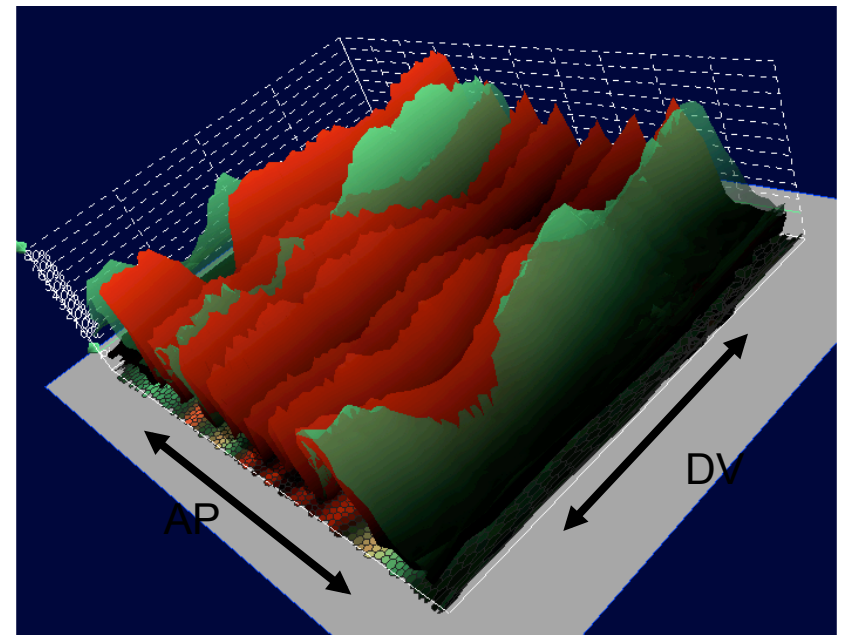


- Entire embryo in one cylindrical projection
- Third dimension can be used for display of additional information

3D Offset Surfaces



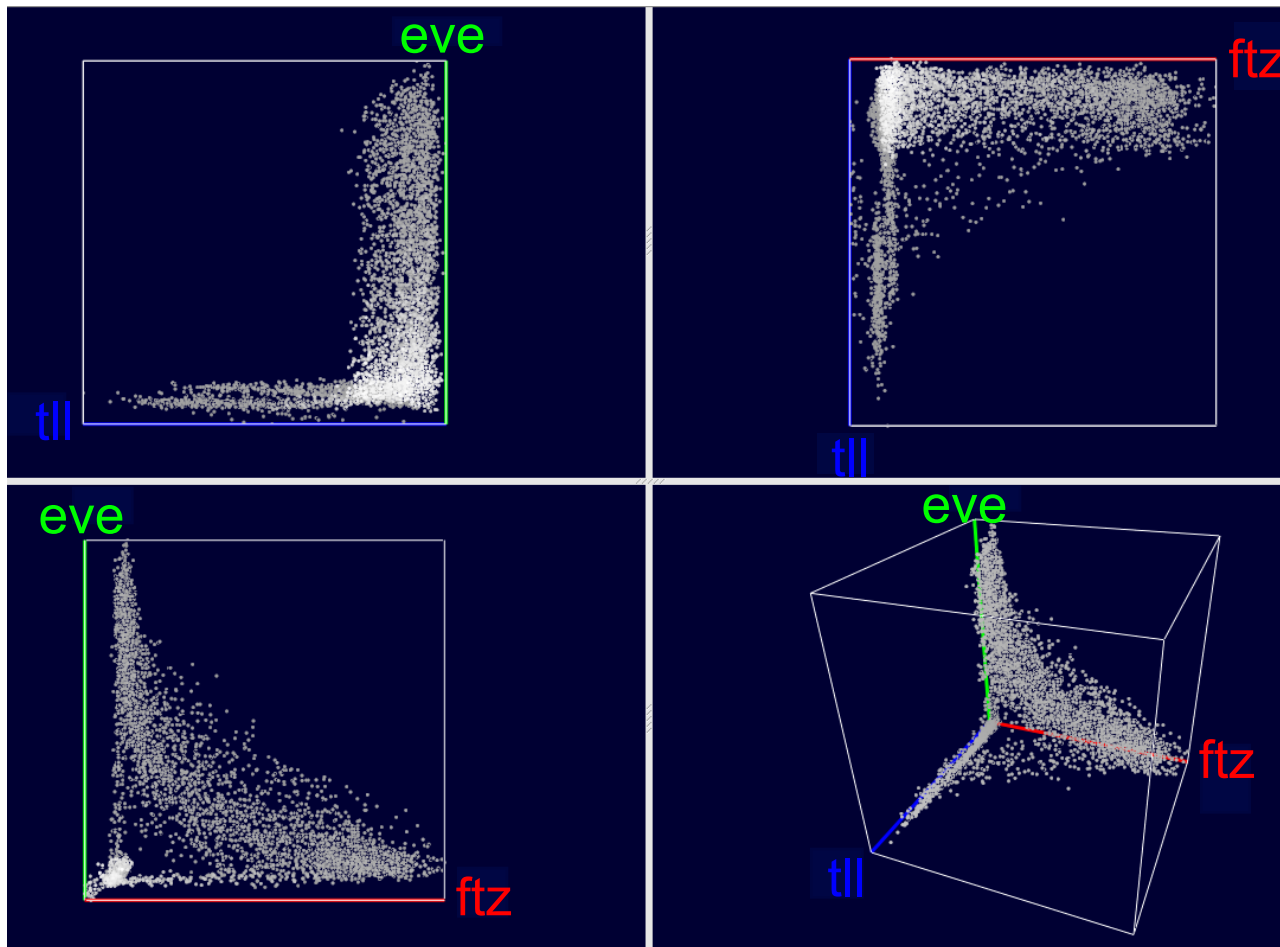
ftz eve



eve hb

- Small differences in expression visible
- Quantitative analysis of gene expression

2D/3D Scatter Plots

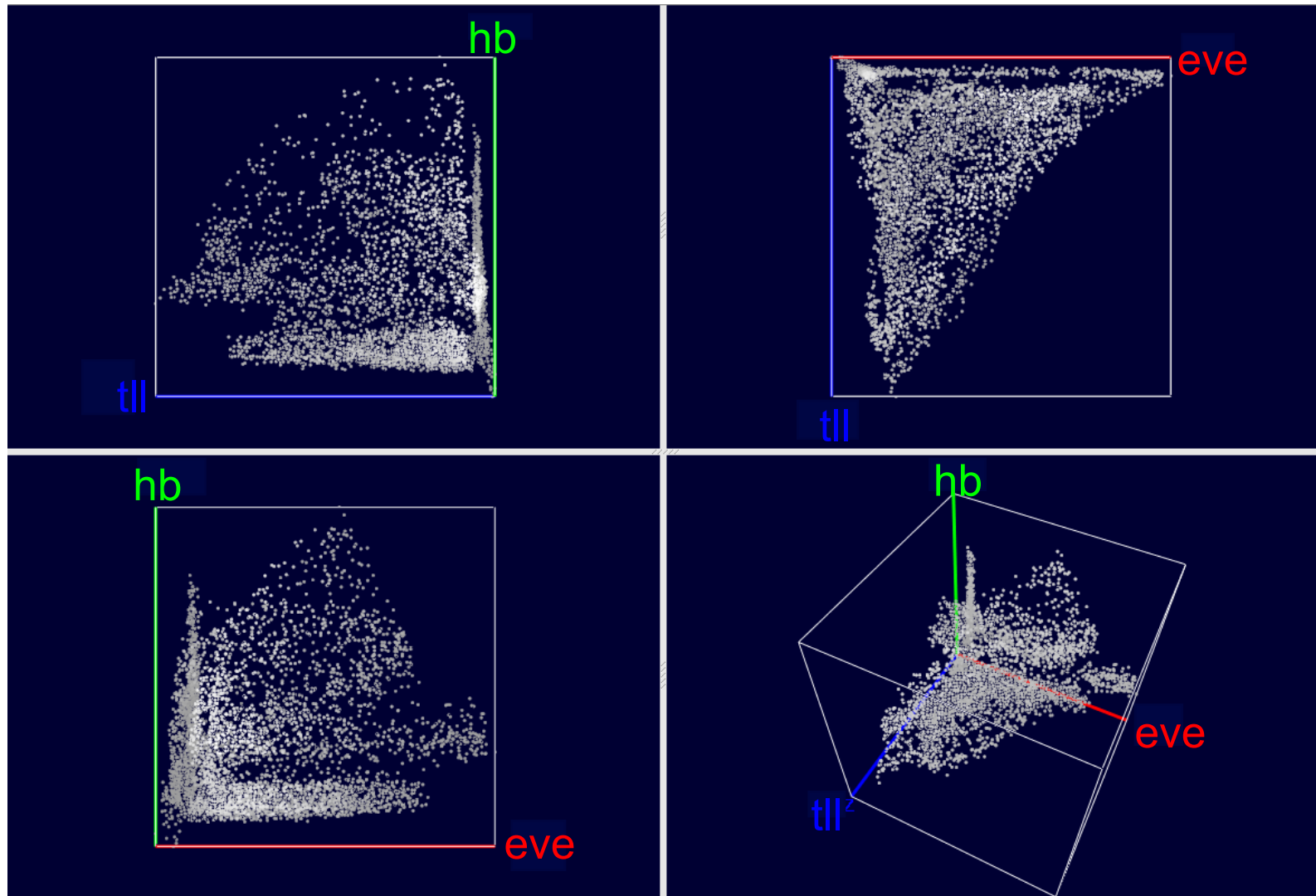


- Identify relationships between gene expression levels

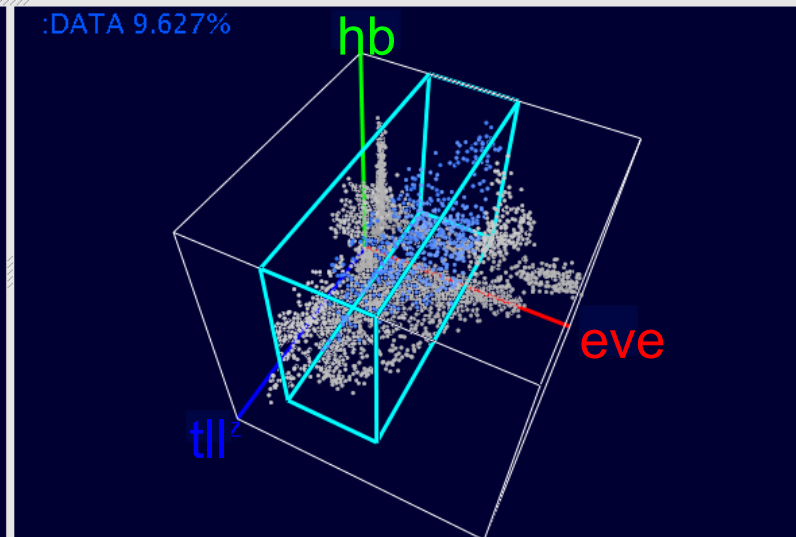
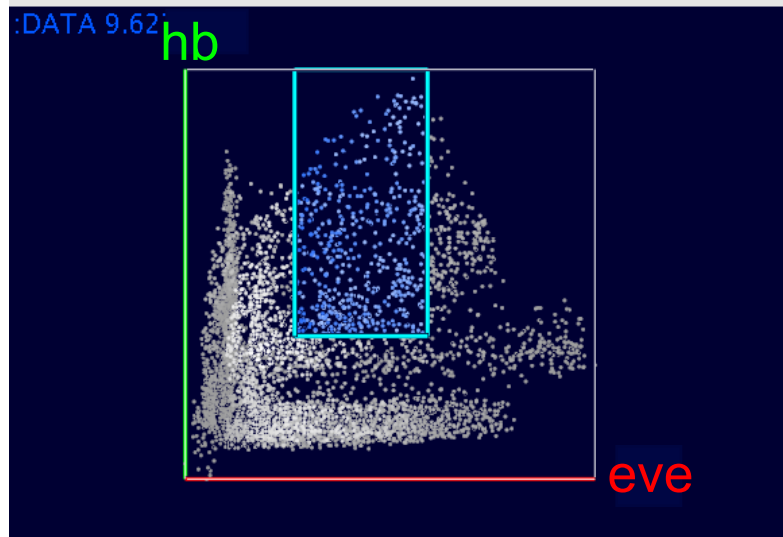
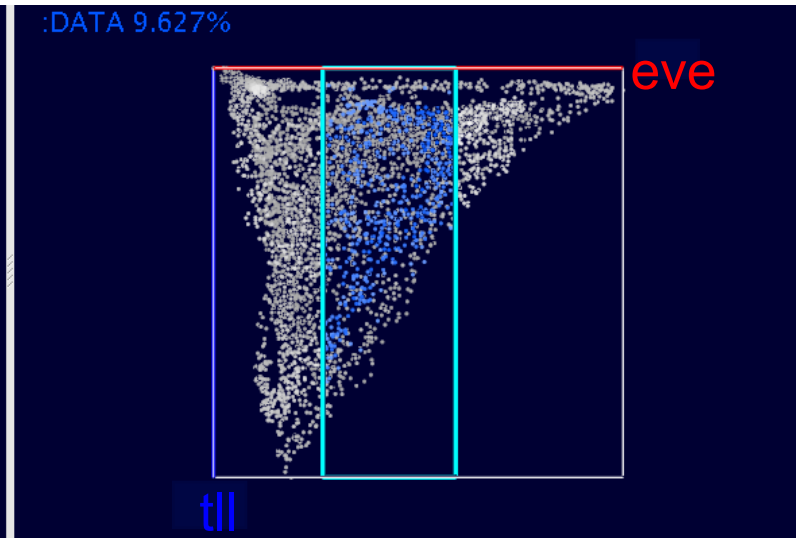
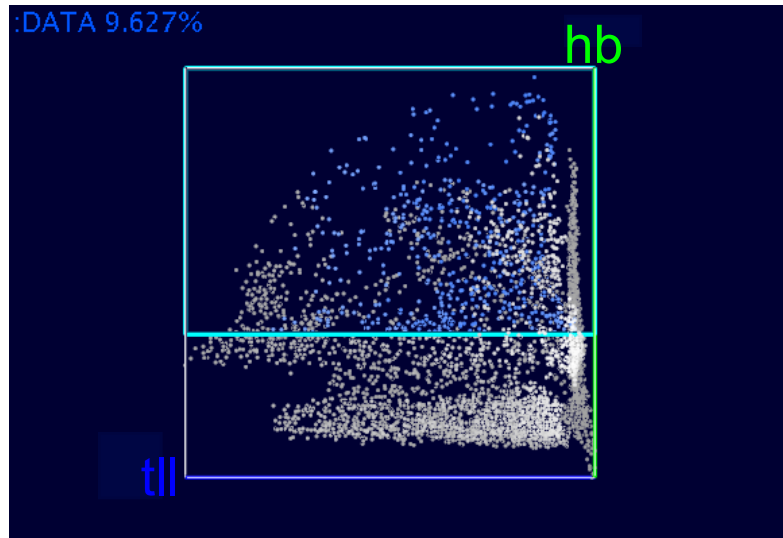
Correlate Views vis “Show & Brush”

- *Brushing*: Select a subset of cells in one view
 - By “painting” on embryo in physical view
 - By choosing expression level ranges in expression space views
- *Linking*: Highlight selected cells in all other views
- **Fundamental Idea:**
 - Select “interesting expression properties” in one view and examine properties of this subset in other views.

Brushing and Linking Example – Scatterplot

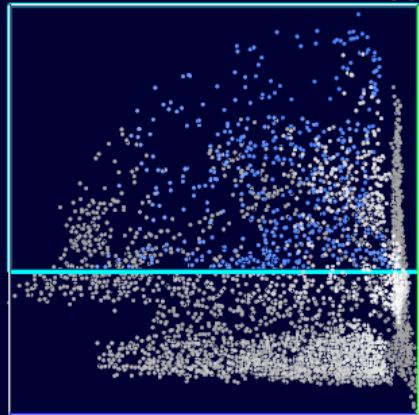


Brushing and Linking Example – Brushing

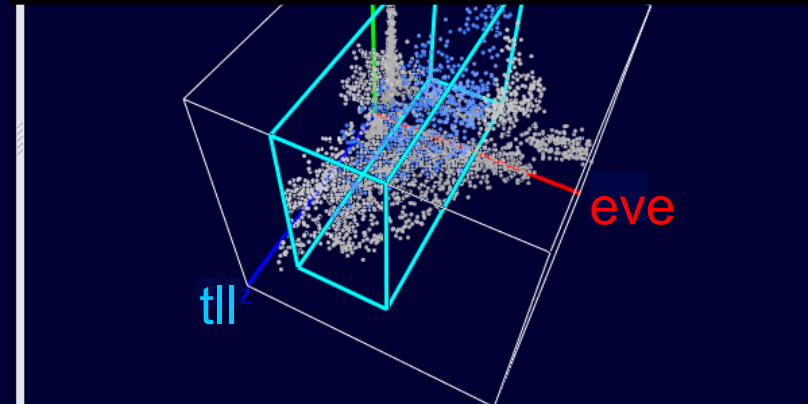
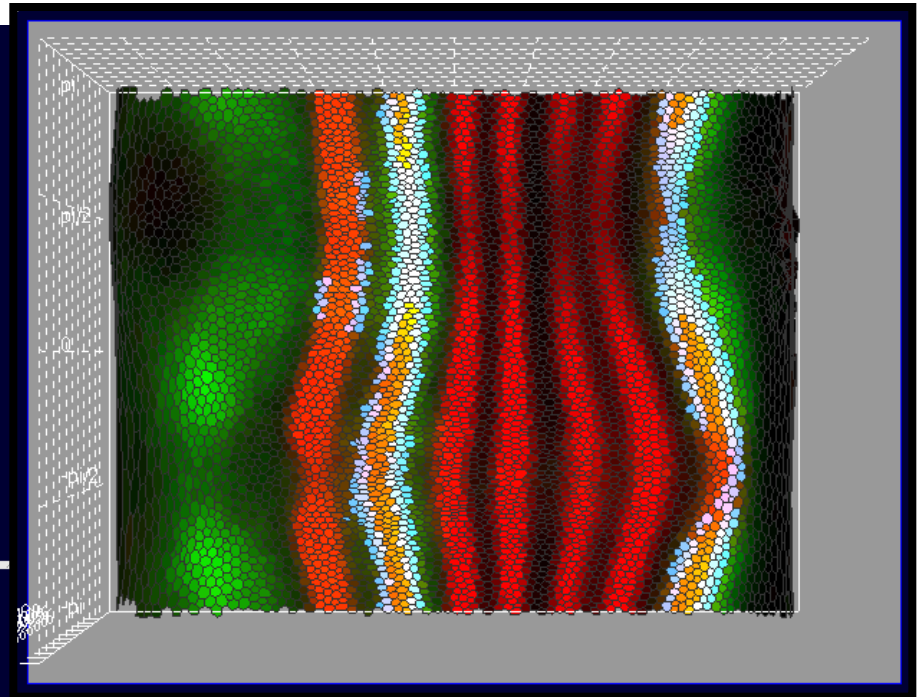
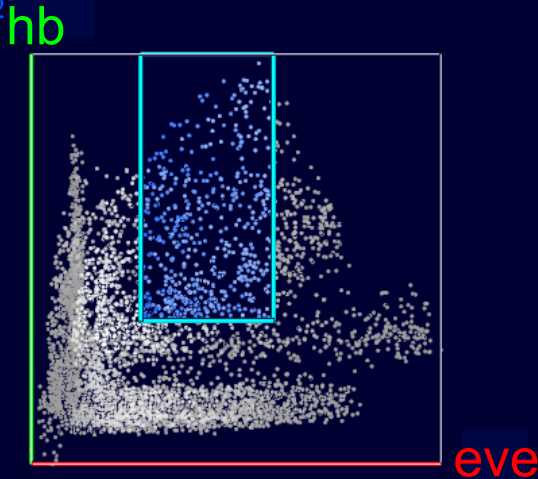


Brushing and Linking Example – Linking

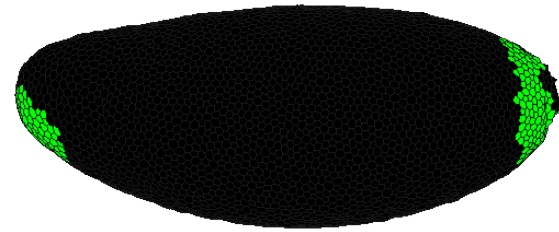
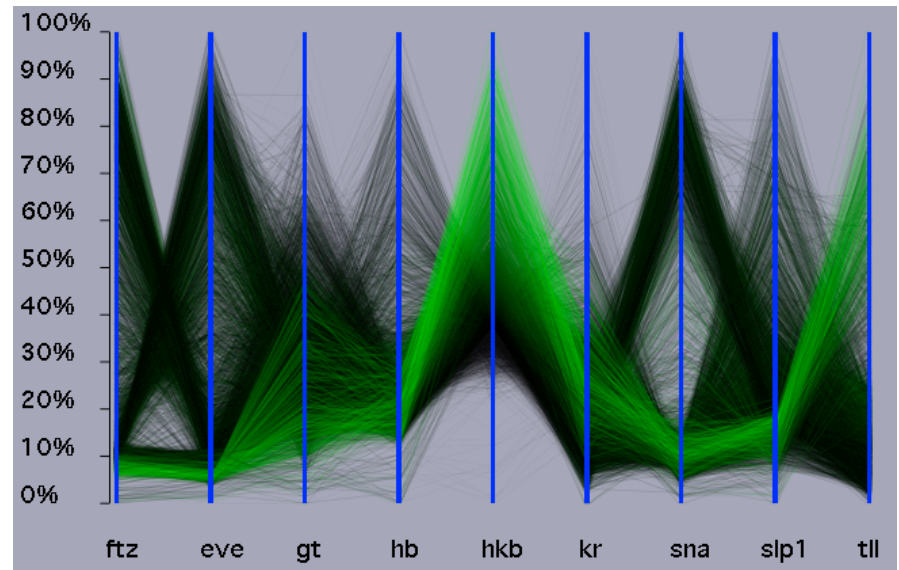
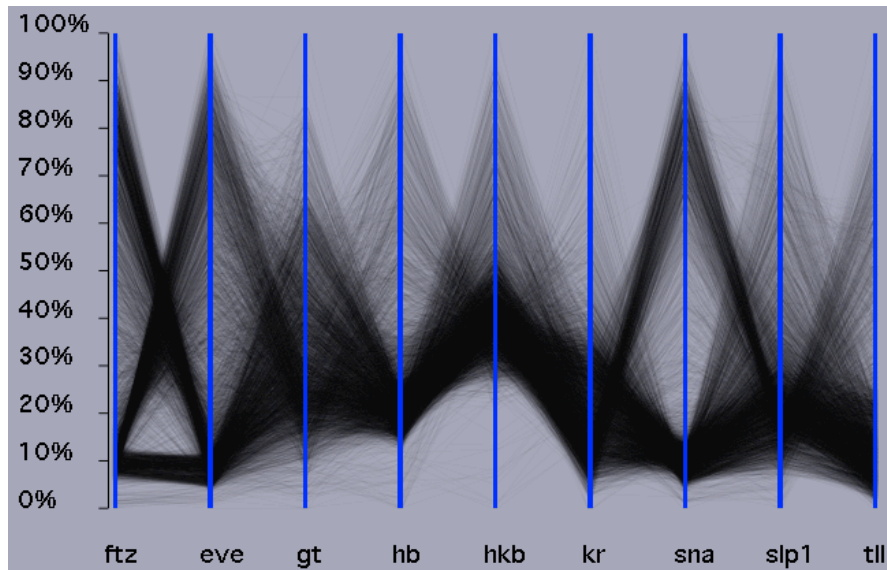
:DATA 9.627%



:DATA 9.62%



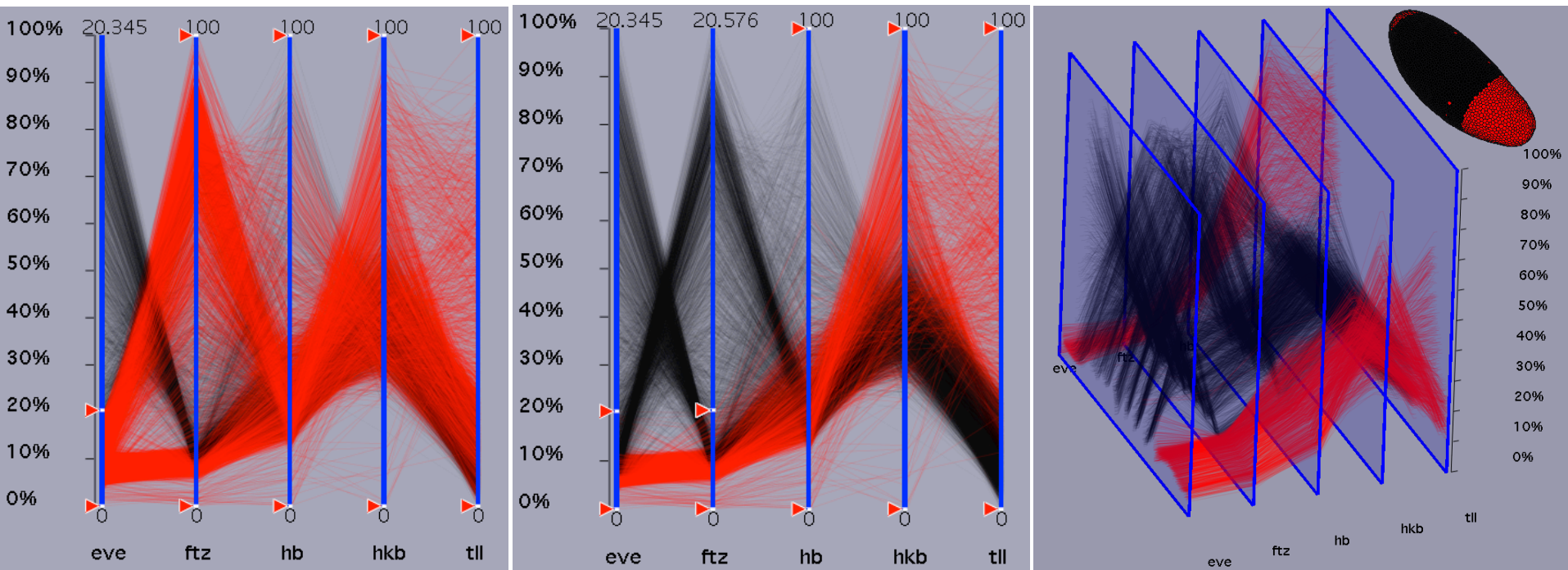
Parallel Coordinates



- Support analysis of relationship of more than three genes

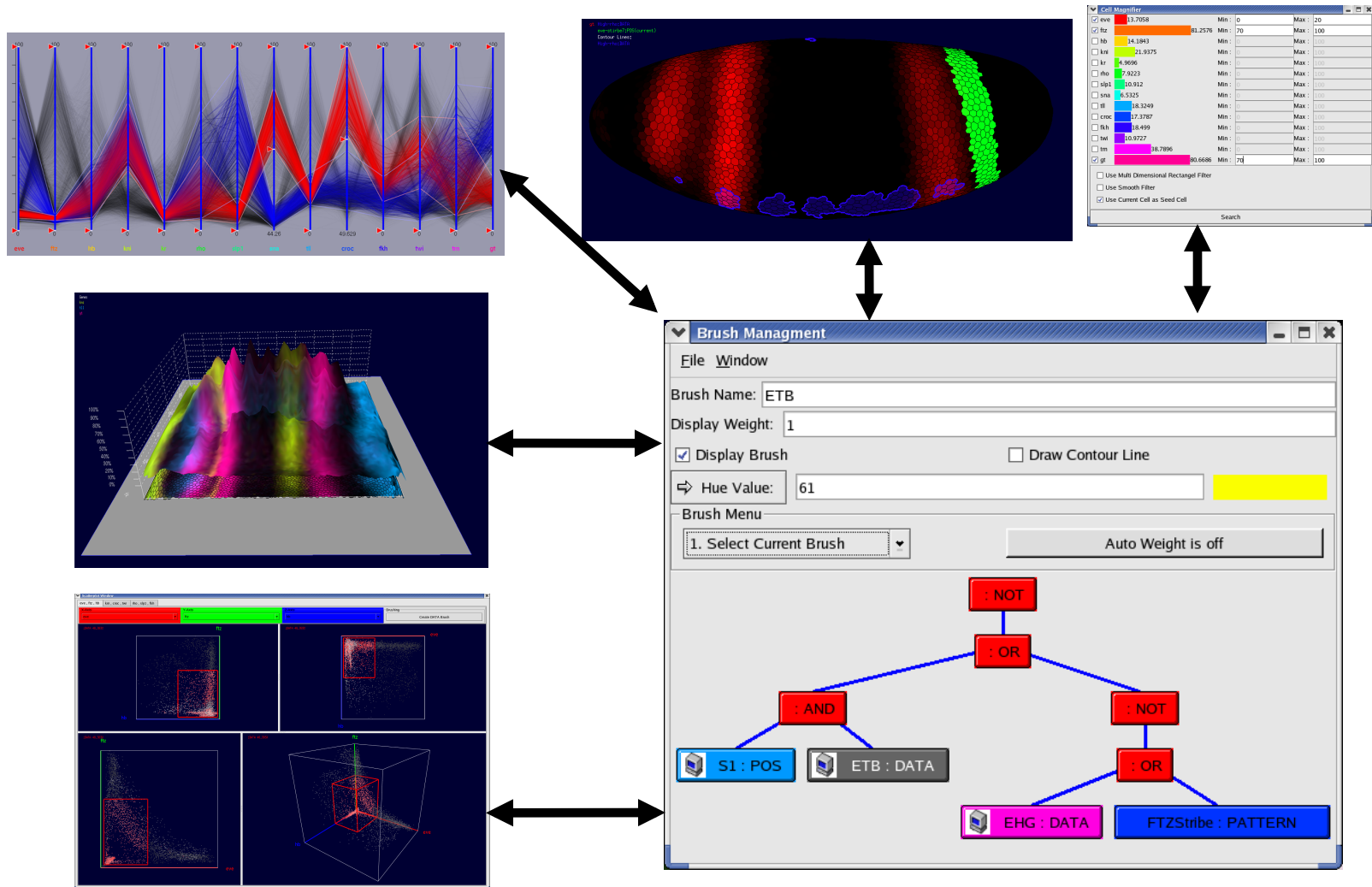
[Rübel et al., Eurographics/IEEE VGTC EuroVis Conference, Lisbon, Portugal, 2006]

Brushing in Parallel Coordinates



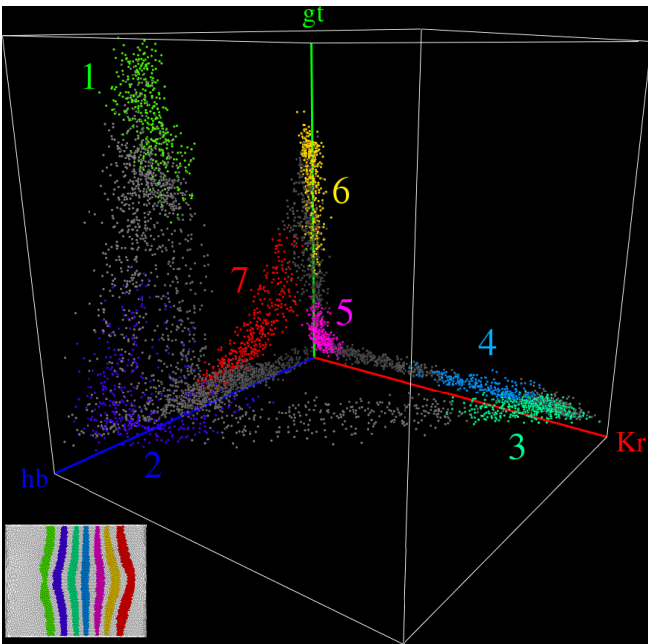
- Selection of expression range along each parallel coordinate axis possible

Linked Views – Combining Selections

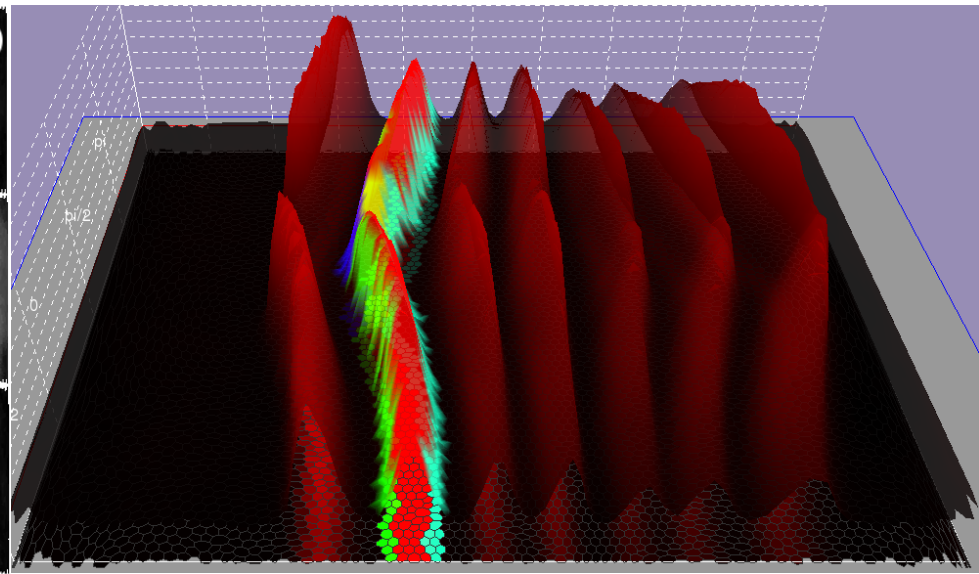
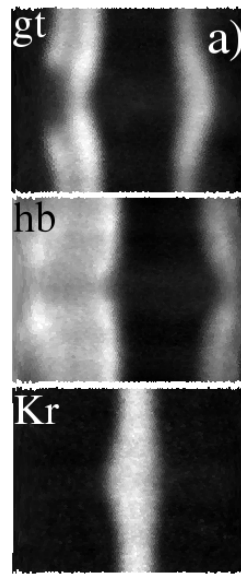


[Weber et al., IEEE TCBB, 2009]

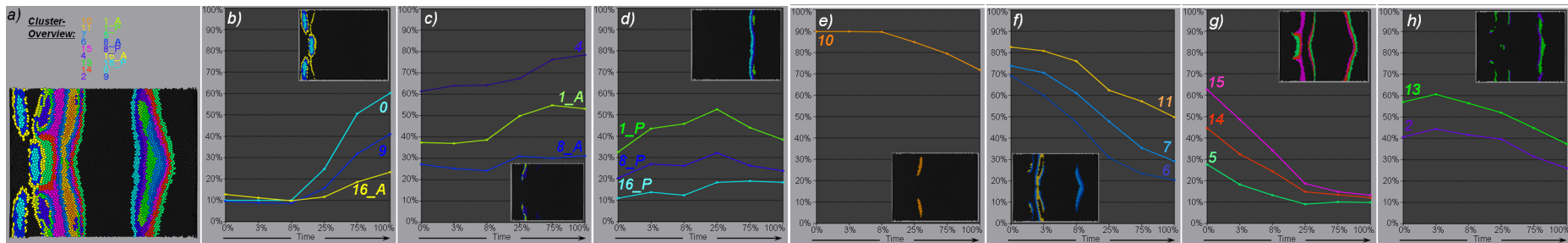
Clustering of 3D Gene Expression



How does the expression pattern of a single gene vary?



How do gene regulatory networks define the expression pattern of a gene?



What is the temporal variation of a gene expression pattern?

[Rübel et al., IEEE TCBB, 2009]

Analysis of ChIP-chip Data

M.-Y. Huang, G.H. Weber, X.-Y. Li, M.D. Biggin, and B. Hamann. *Quantitative Visualization of ChIP-chip Data by Using Linked Views.*

In: Proc. IEEE BIBM 2010 Workshops, Workshop on Integrative Data Analysis in Systems Biology (IDASB), pp. 195-200. (2010)

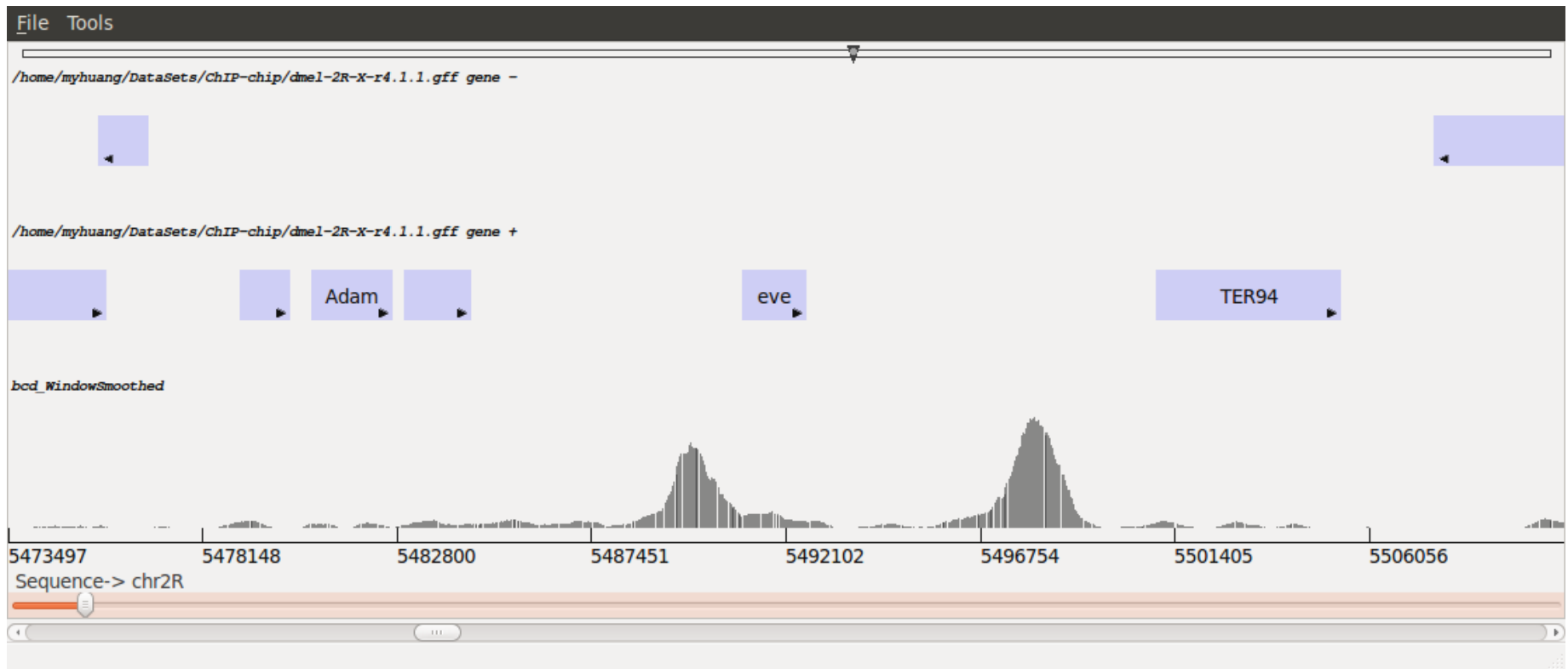


Motivation

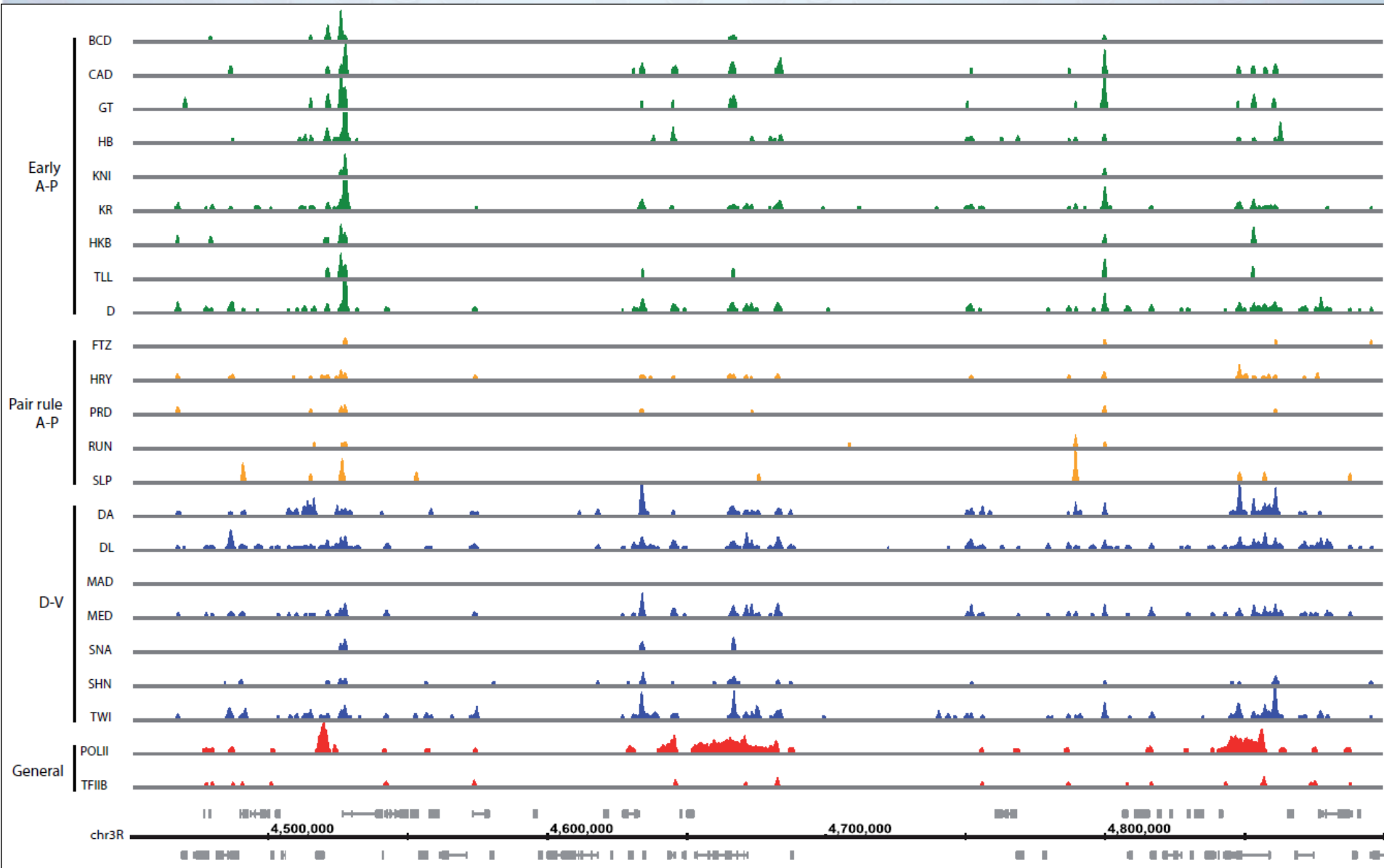
- Most analyses of ChIP-chip *in vivo* binding focused on qualitative description: region bound by certain factor or not
- Increasing evidence suggests that
 - Factors bind in highly overlapping manner on same genomic regions
 - Quantitative differences critical determinants of different biological specificity of factors
- Need tool to facilitate quantitative visualization of differences between transcription factors and genomic regions they bind to understand each factor's unique roles in the network
- Build this tool based on IVA
- Interesting example since no “physical” / 3D views

ChIP-chip Data

- ChIP-chip data describe interaction between protein and DNA *in vivo*, e.g., location of binding



ChIP-chip Data

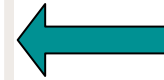
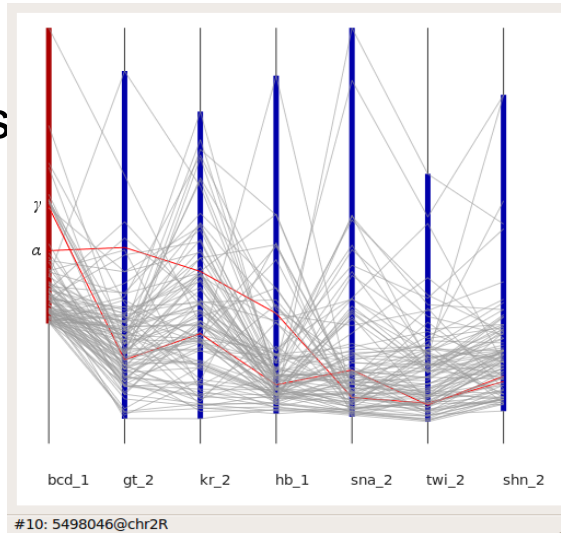


Input and Data Processing

- **Input:**
 - ChIP-chip data
 - Peak locations
- **Processing:**
 - Data smoothing
 - Correlation computation
 - i. Choose a transcription factor (TF), fa , as the base
 - ii. Pick a subset of its primary peaks
 - iii. Look up corresponding ChIP-chip scores of other TFs at these peak locations
 - iv. Pick another TF as the new base and repeat step i~iii

Overview

Parallel Coordinates



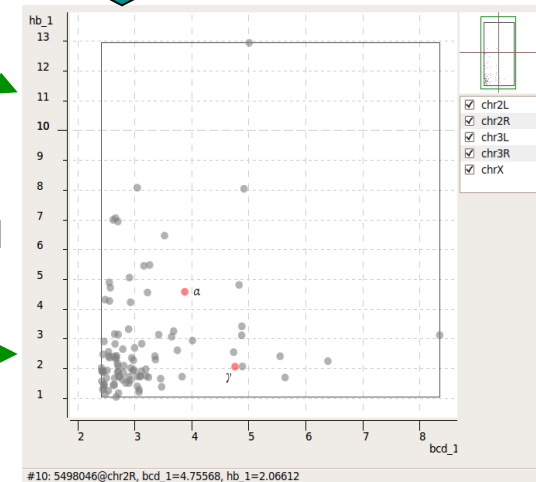
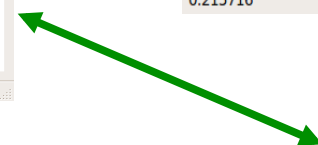
Transcription Factor Option

	bcd_1	gt_2	kr_2	hb_1	sna_2	twi_2	shn_2
bcd_1	1.00	0.25	0.25	0.22	-0.10	-0.15	-0.01
gt_2	0.24	1.00	0.36	0.04	-0.05	-0.13	-0.01
kr_2	0.31	0.33	1.00	-0.03	0.03	0.22	0.01
hb_1	0.45	0.38	0.48	1.00	0.52	0.30	0.30
sna_2	0.11	-0.02	-0.09	0.49	1.00	0.22	0.22
twi_2	0.12	0.16	0.10	0.32	0.14	1.00	0.22
shn_2	0.14	-0.04	-0.03	0.32	0.14	0.14	1.00

0.215716

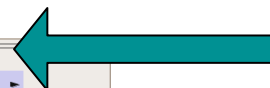
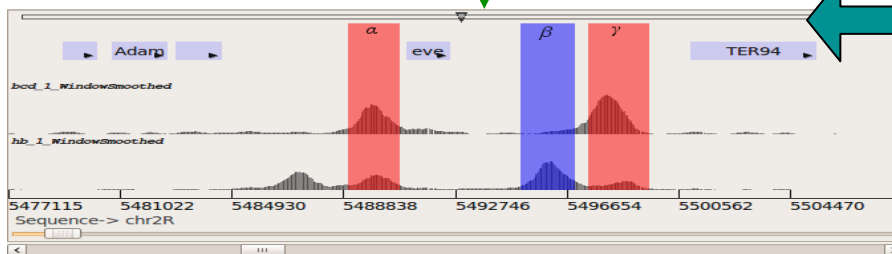
Color scale: ≥ 0.60 , 0.30, 0.00, -0.30, ≤ -0.60

Correlation Table



Scatter Plots

Track View

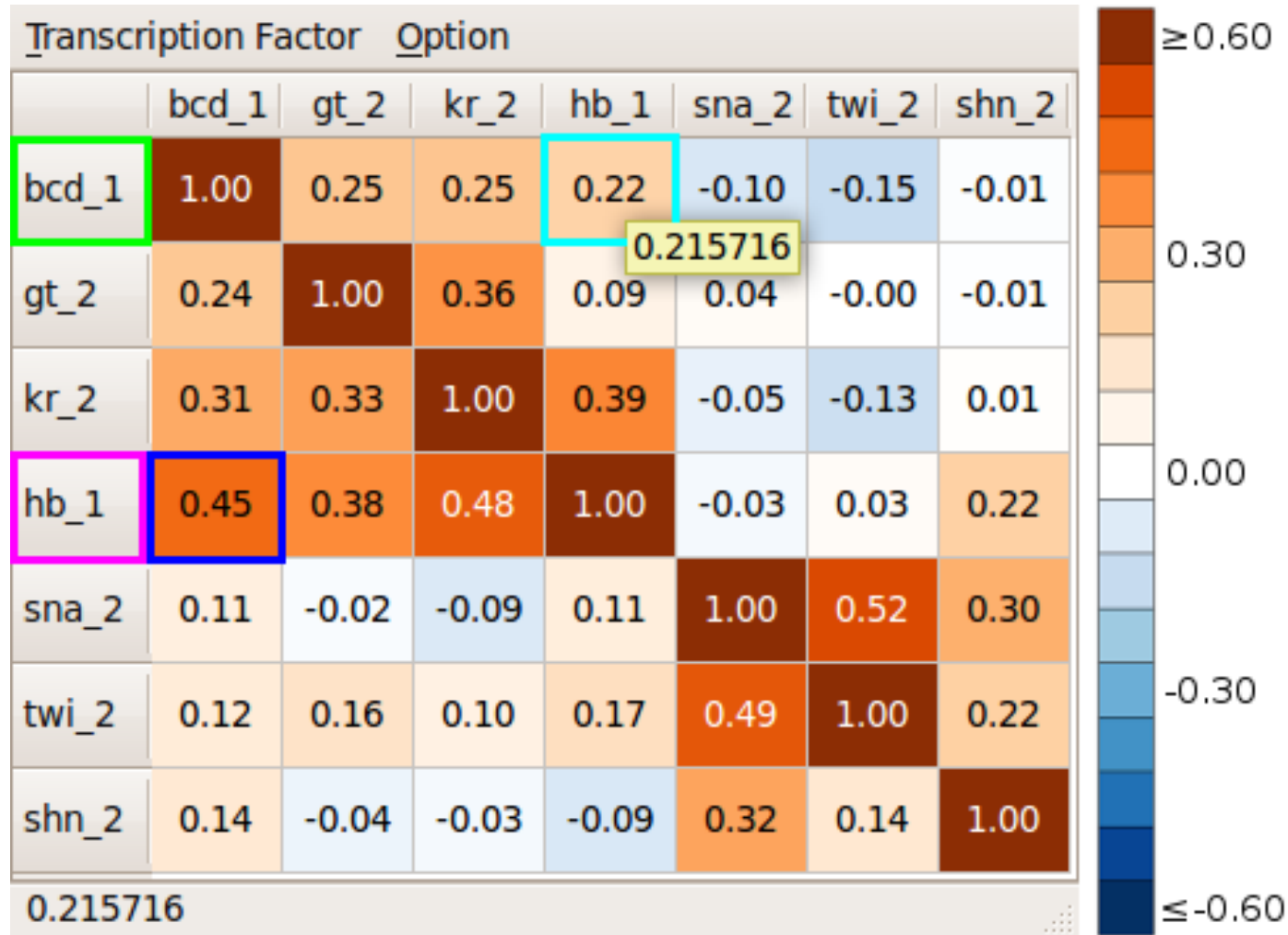


View creation

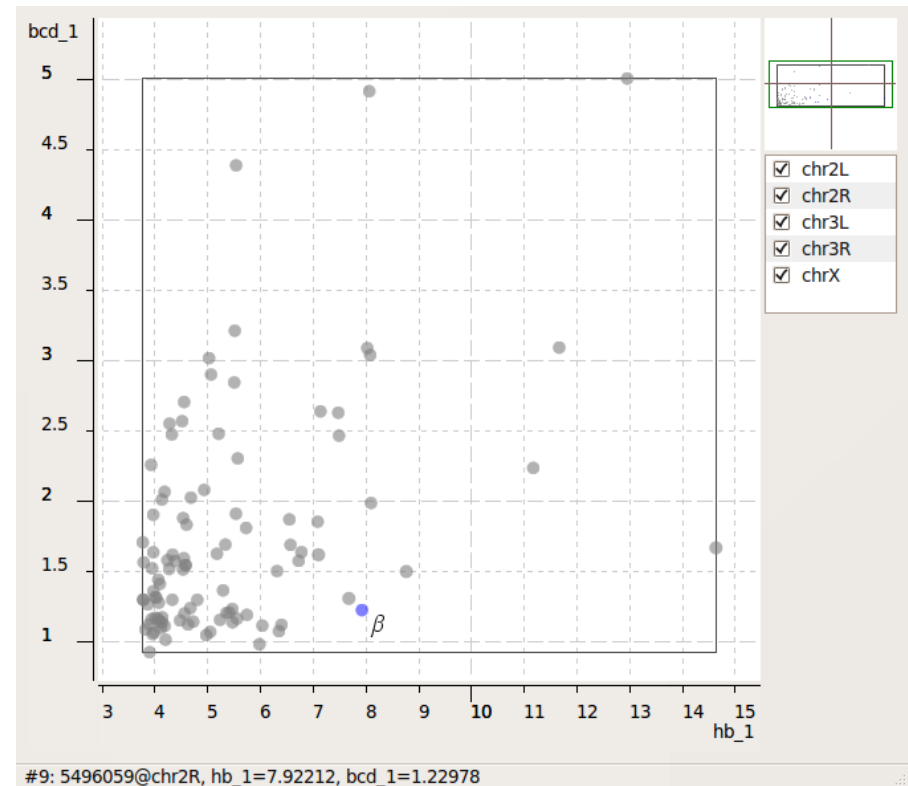
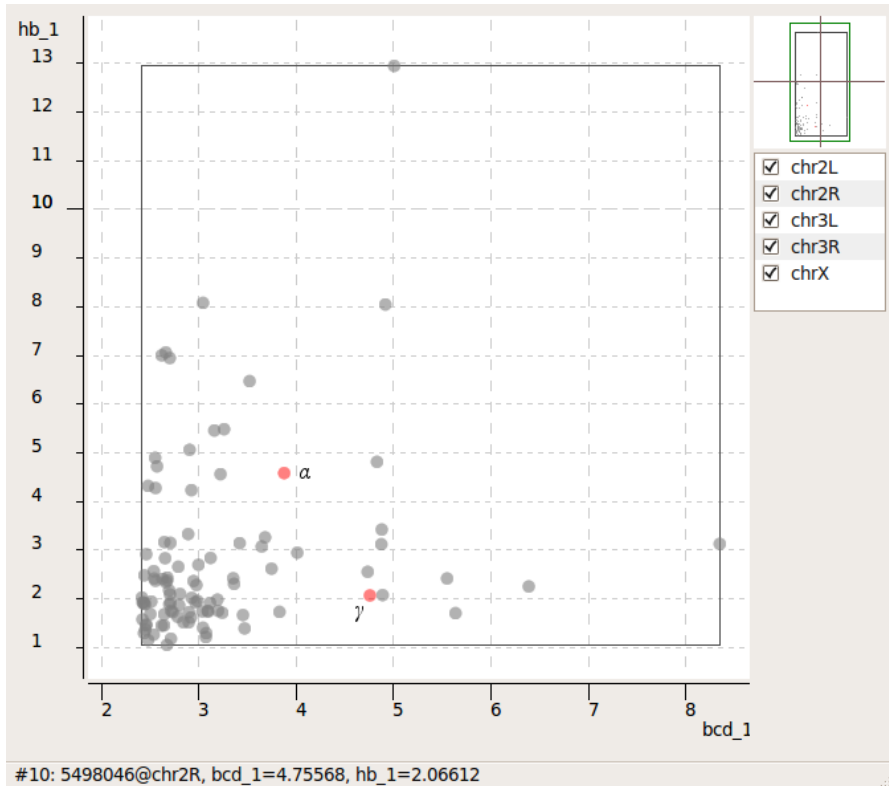


Brushing-and-Linking

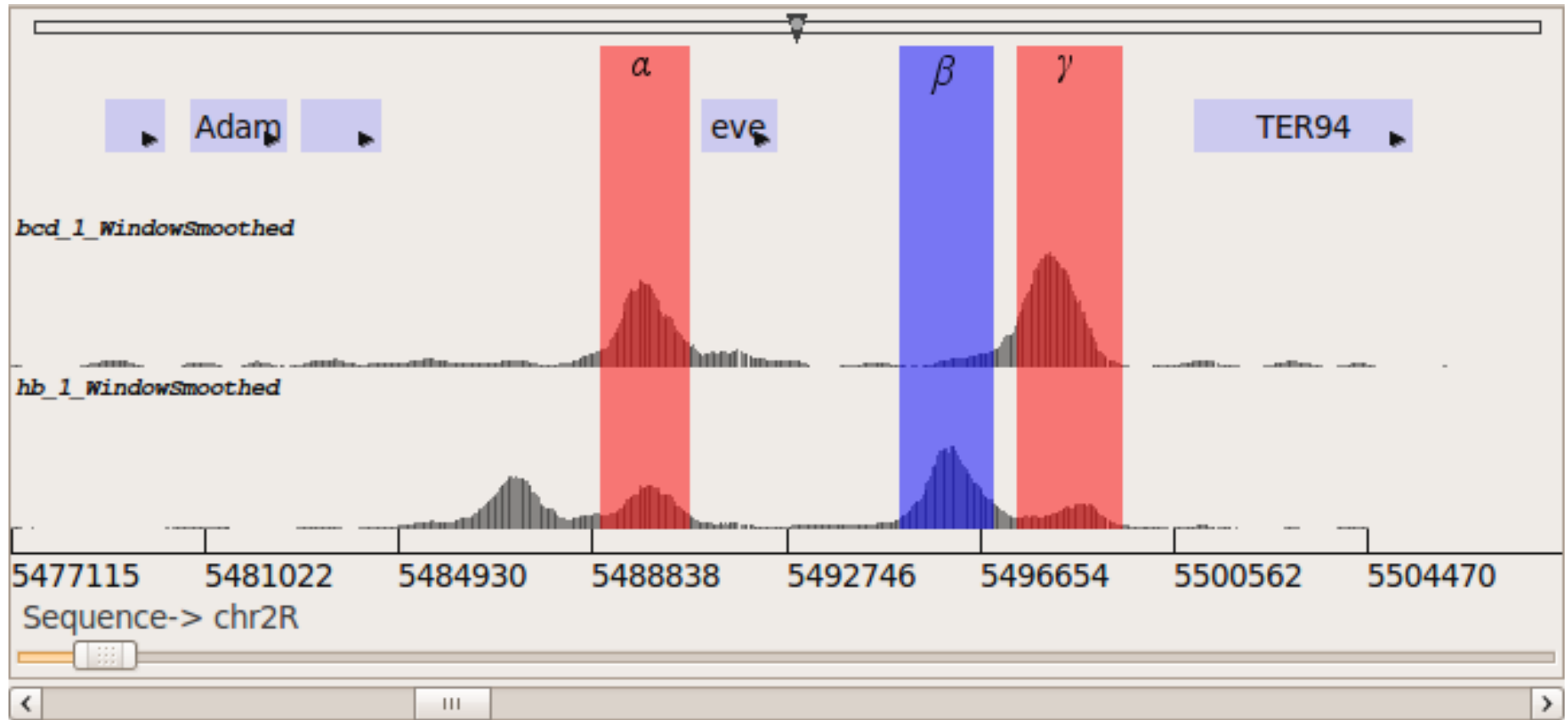
Correlation table



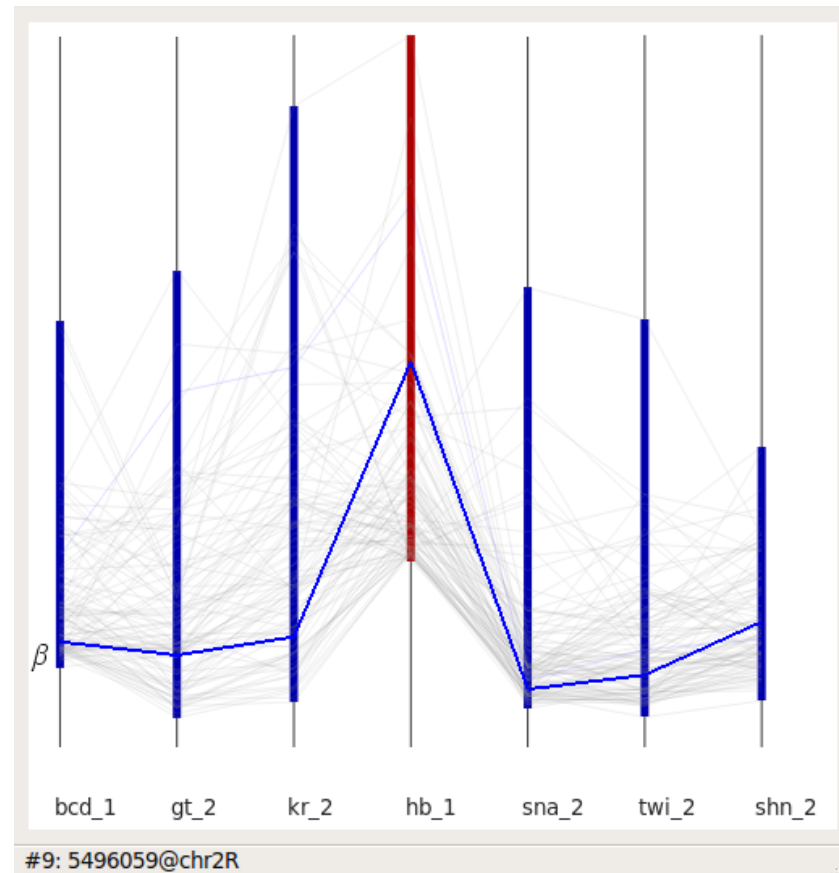
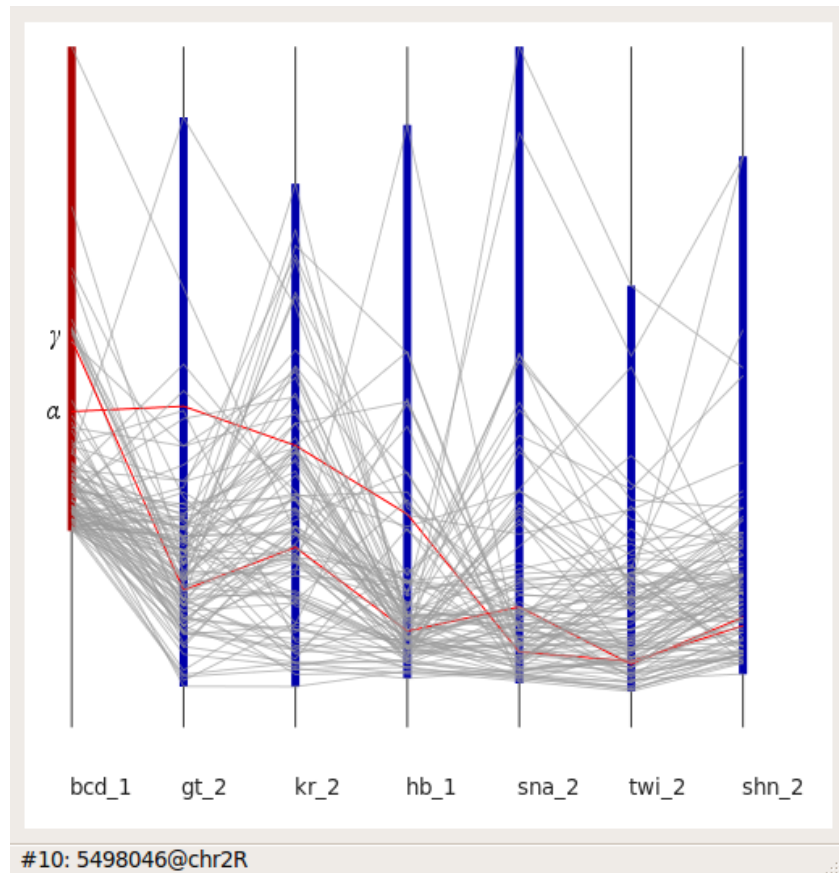
Scatter Plots



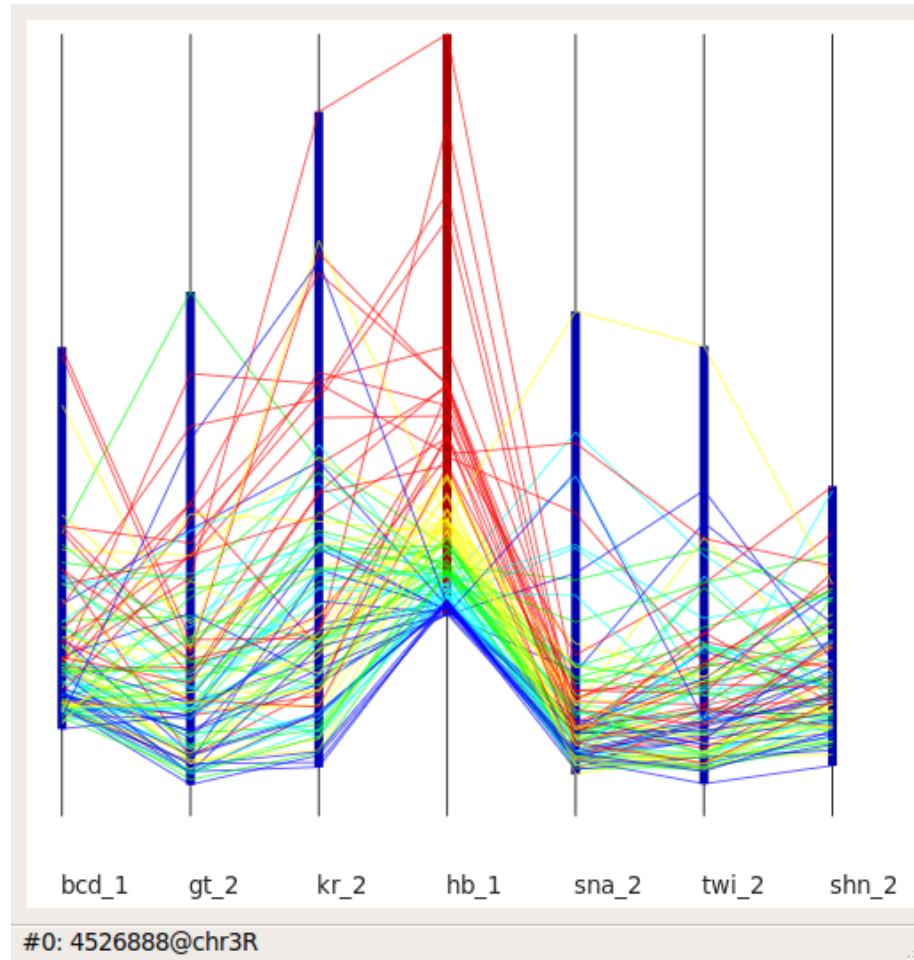
Track View



Parallel Coordinates

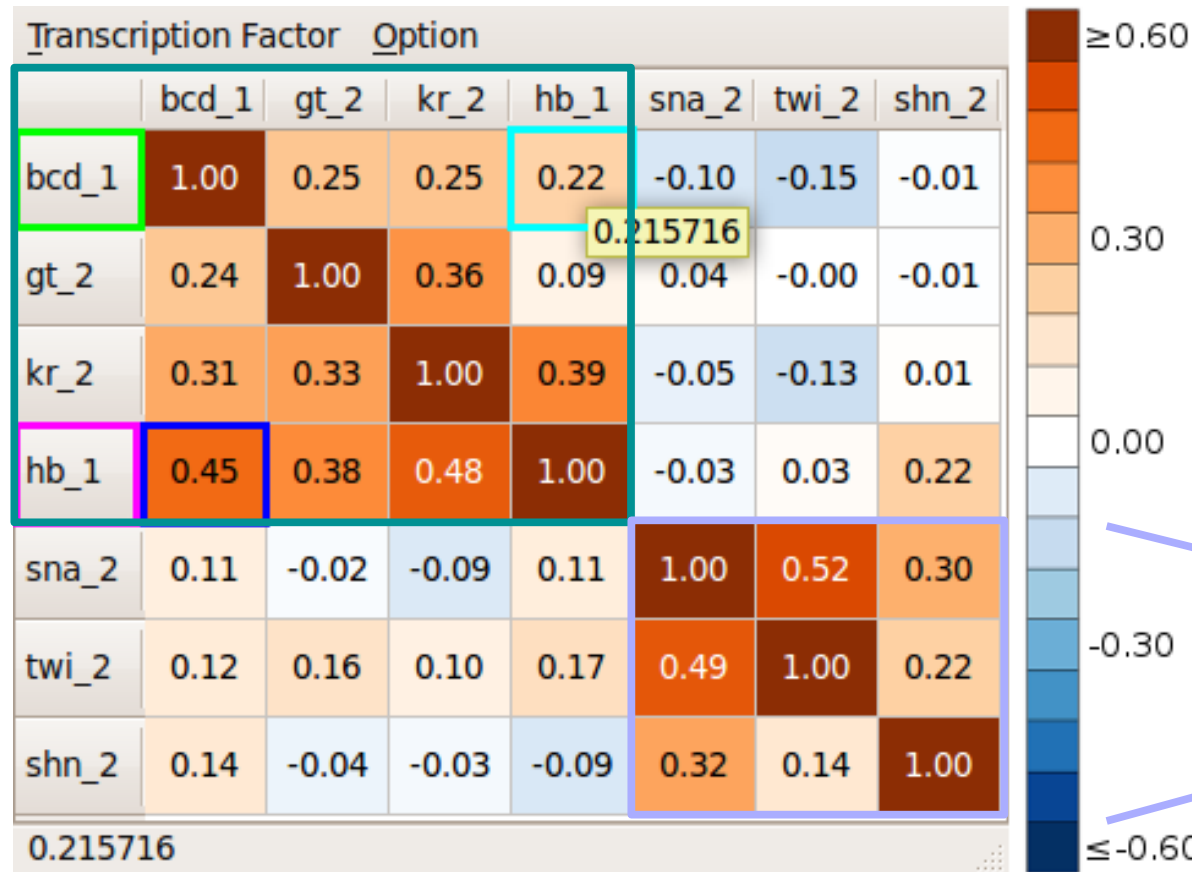


Parallel Coordinates



Case Study (1/5)

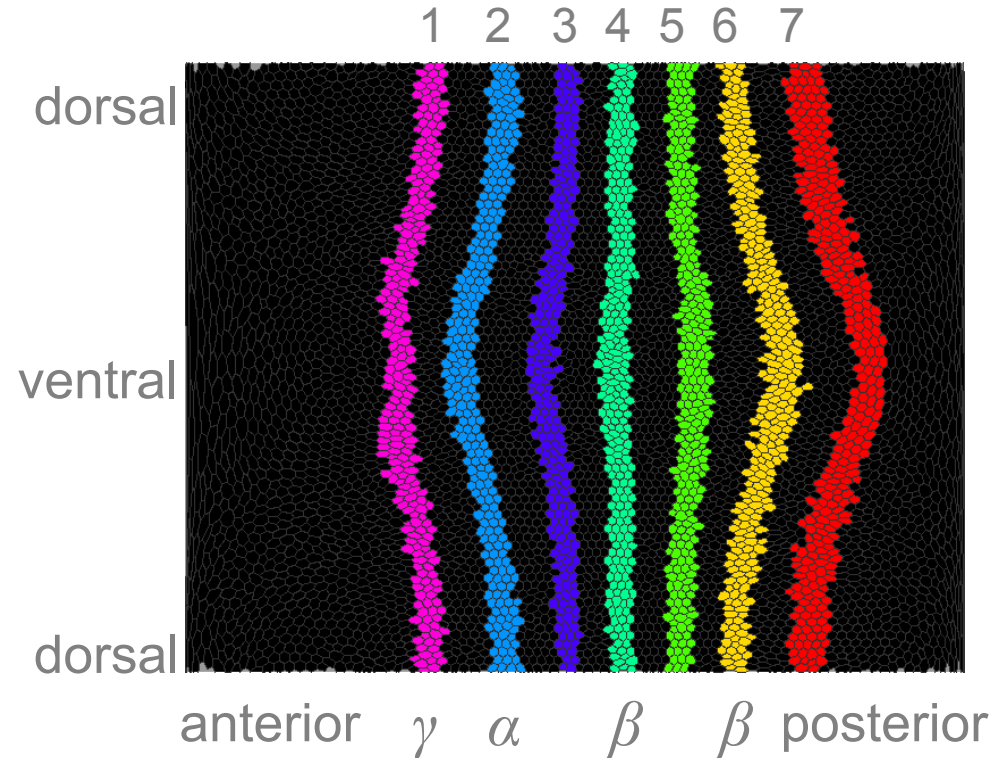
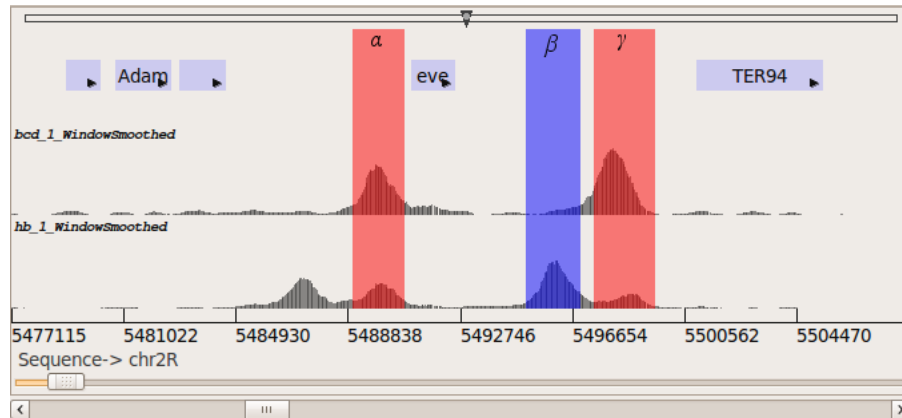
A-P Factors



D-V Factors

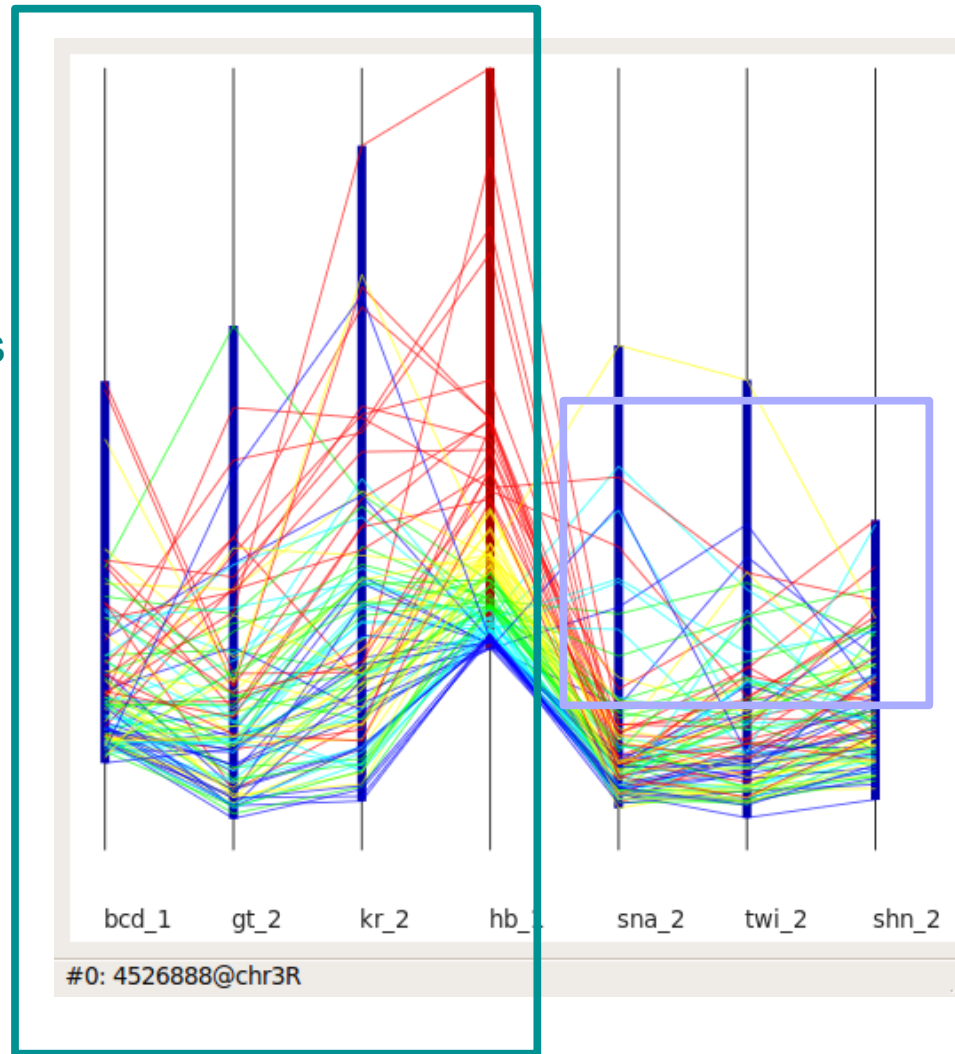
Case Study (2/5)

- *Eve*
 - α : stripe 2, *bcd* & *hb* activate
 - β : stripe 4/6, *hb* represses
 - γ : stripe 1, *bcd* activates



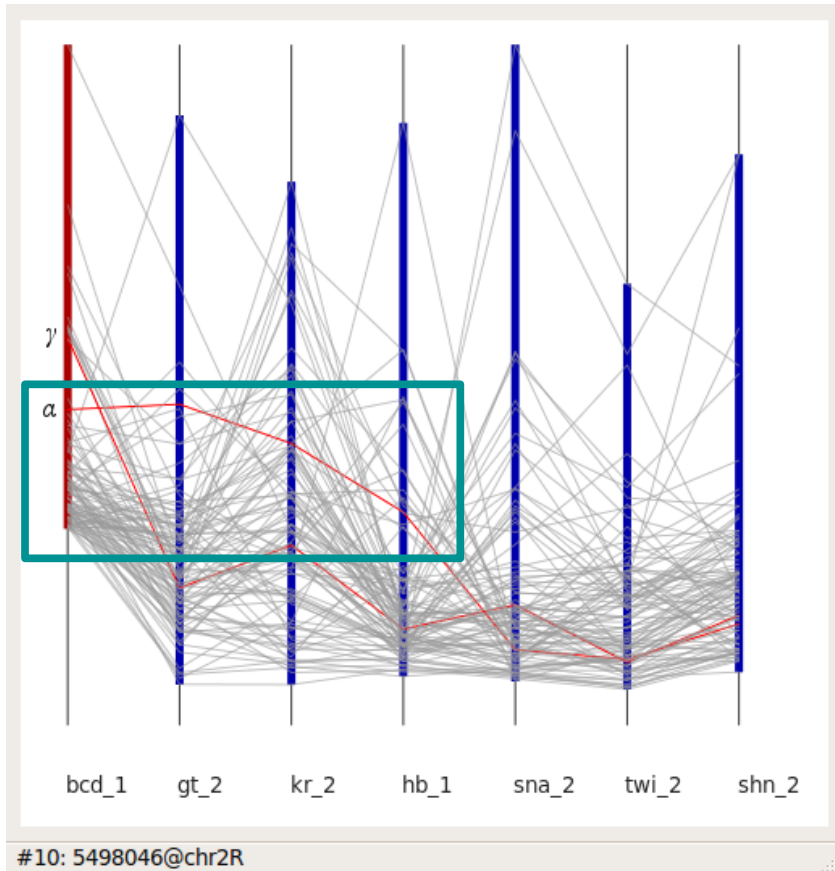
Case Study (3/5)

Higher correlation
with other A-P factors



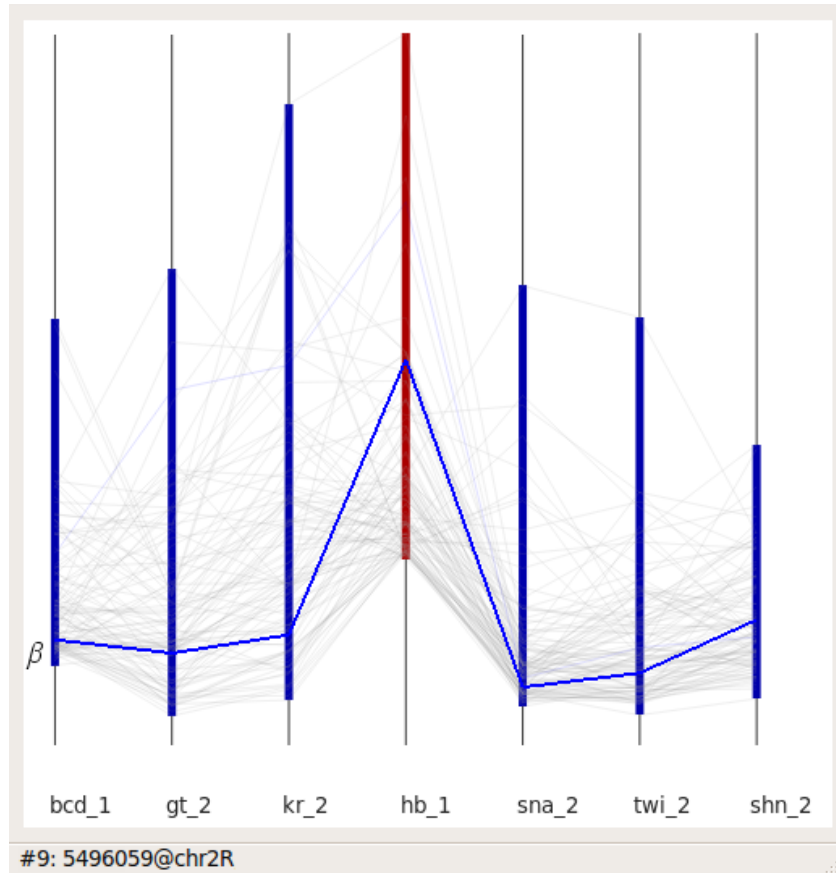
Still have regions
bound to high *hb* and
moderate D-V factors

Case Study (4/5)



- α is bound strongly not only by *bcd* and *hb*, but also by *kr* and *gt* (two factors known to be important for defining the expression boundaries of *eve* stripe 2)
- γ (*eve* stripe 1 enhancer) is only strongly bound by *bcd*

Case Study (5/5)



- β (*eve* stripe 4/6) is only strongly bound by *hb*

Literature

- O. Rübel, S.V.E. Keränen, M.D. Biggin, D.W. Knowles, G.H. Weber, H. Hagen, B. Hamann, and E.W. Bethel, "Linking Advanced Visualization and MATLAB for the Analysis of 3D Gene Expression Data," Series: Mathematics and Visualization, Visualization in Medicine and Life Sciences II, Progress and New Challenges, Springer Verlag, Heidelberg, Germany, 2012, pp 267-285. LBNL-4891E.
- Min-Yu Huang, Gunther H. Weber, Xiao-Yong Li, Mark D. Biggin, and Bernd Hamann. "Quantitative Visualization of ChIP-chip Data by Using Linked Views." In: Proceedings IEEE International Conference on Bioinformatics and Biomedicine 2010 (IEEE BIBM 2010) Workshops, Workshop on Integrative Data Analysis in Systems Biology (IDASB), pp. 195-200. LBNL-4491E.
- O. Rübel, G. H. Weber, M-Y Huang, E. W. Bethel, M. D. Biggin, C. C. Fowlkes, C. Luengo Hendriks, S. V. E. Keränen, M. Eisen, D. Knowles, J. Malik, H. Hagen and B. Hamann, "Integrating Data Clustering and Visualization for the Analysis of 3D Gene Expression Data." IEEE Transactions on Computational Biology and Bioinformatics, Vol 7, No.1, Pages 64-79, Jan/Mar 2010. LBNL-382E.
- G. H. Weber, O. Rübel, M.-Y. Huang, A. H. DePace, C. C. Fowlkes, S. V. E. Keränen, C. L. Luengo Hendriks, H. Hagen, D. W. Knowles, J. Malik, M. D. Biggin and B. Hamann. "Visual exploration of three-dimensional gene expression using physical views and linked abstract views." In *IEEE Transactions on Computational Biology and Bioinformatics.*, 6(2), April-June, pp. 296-309, 2009. LBNL-63776.
- O. Rübel, G.H. Weber, S.V.E. Keränen, C.C. Fowlkes, C.L. Luengo Hendriks, L. Simirenko, N.Y. Shah, M.B. Eisen, M.D. Biggin, H. Hagen, J.D. Sudar, J. Malik, D.W. Knowles, and B. Hamann. PointCloudXplore: Visual analysis of 3D gene expression data using physical views and parallel coordinates. In: B. Sousa Santos, T. Ertl, and K.I. Joy, eds., *Data Visualization 2006 (Proceedings of EuroVis 2006)*, Eurographics Association, Aire-la-Ville, Switzerland, pp. 203-210. May, 2006. LBNL-60005.